Quantification and depth distribution analysis of carbon to nitrogen ratio in forest soils using reflectance spectroscopy

Asa Gholizadeh ^a, Mohammadmehdi Saberioon ^b, Nastaran Pouladi ^a, Eyal Ben-Dor ^c

^a Department of Soil Science and Soil Protection, Faculty of Agrobiological Sciences, Food and Natural Resources, Czech University of Life Sciences Prague, 16500, Prague, Czech Republic

^b Helmholtz Centre Potsdam GFZ German Research Centre for Geosciences, Section 1.4 Remote Sensing and Geoinformatics, Telegrafenberg, Potsdam 14473, Germany

^c Remote Sensing Laboratory, Department of Geography and Human Environment, Porter School of Environment and Earth Science, Tel Aviv University, Tel Aviv 69978, Israel

Abstract

Forest soils have large contents of carbon (C) and nitrogen (N), which have significant spatial variability laterally across landscapes and vertically with depth due to decomposition, erosion and leaching. Therefore, the ratio of C to N contents (C:N), a crucial indicator of soil quality and health, is also different depending on soil horizon. These attributes can cost-effectively and rapidly be estimated using visible–near infrared–shortwave infrared (VNIR–SWIR) spectroscopy. Nevertheless, the effect of different soil layers, particularly over large scales of highly heterogeneous forest soils, on the performance of the technique has rarely been attempted. This study evaluated the potential of VNIR–SWIR spectroscopy in quantification and variability analysis of C:N in soils from different organic and mineral layers of forested sites of the Czech Republic. At each site, we collected samples from the litter (L), fragmented (F) and Humus (H) organic layers, and from the A₁ (depth of 2–10 cm) and A₂ (depth of 10–40 cm) mineral layers providing a total of 2505 samples. Support vector machine regression (SVMR) was used to train the prediction models of the selected attributes at each individual soil layer and the merged layer (profile). We further produced the spatial distribution maps of C:N as the target attribute at each soil layer. Results showed that the prediction accuracy based on the profile spectral data was adequate for all attributes. Moreover, F was the most accurately predicted layer, regardless of the soil attribute. C:N models and maps in the organic layers performed well although in mineral layers, models were poor and maps were reliable only in areas with low and moderate C:N. On the other hand, the study indicated that VNIR–SWIR spectroscopy could efficiently predict and map organic layers of the Czech Republic's forested sites but in mineral layers, it didn't differentiate classes of C:N higher than 50.

Keywords: Forest soil, Soil organic carbon, C:N, Soil horizons, VNIR–SWIR spectroscopy.

1. Introduction

Soil organic carbon (SOC) is the largest terrestrial carbon (C) pool in the biosphere (Lal, 2008) and plays a key role in mitigating global atmospheric carbon dioxide (CO₂) level. SOC has several significant functions in the environment (Murphy, 2014) such as maintaining soil structure and physical stability. Accordingly, it is crucial to a number of natural processes linked to soil health

40 and fertility (Dinakaran et al., 2016). Soil nitrogen (N) is also one of the important indicators of
41 the ecological environment and part of the organic matter components and mineralization process
42 in soil (Li et al., 2014; Shang et al., 2014). These attributes affect the supply of food, air, fresh
43 water and agritourism and even associate to the sustainable development of the local economy
44 (Zhang et al., 2007; Keesstra et al., 2012). They have also been used as proxies to assess soil quality
45 and land degradation in some Himalayan and Mediterranean areas (Kukal et al., 2014; Munoz-
46 Rojas et al., 2015).

47 Forest soils contain a large amount of the world's SOC, N and energy resources. They influence
48 the quality and composition of the atmosphere and form climate conditions on different scales
49 (Baciak et al., 2015). In forest ecosystems, SOC and N cycles are related via a number of
50 fundamental processes including litterfall, primary production, mineralization of organic matter
51 components and decomposition rate (Alberchtova et al., 2008). The ratio of SOC to N (hereafter
52 C:N) in soil, particularly in forest soil, is another important indicator of soil fertility and quality
53 reflecting the interaction or coupling between SOC and N (Lou et al., 2012; Xu et al., 2018), which
54 is considered as a proxy of C sequestration potential in soils (Akselsson et al., 2005; Vries et al.,
55 2006). Soil C:N also delivers information about the decomposition stage of soil organic matter that
56 is known as an essential factor of soil fertility, and hence highlights the microorganism activities
57 in soil. High values of C:N (> 40) demonstrate the availability of fresh organic matter, whereas
58 lower values (around 5–15) indicate partially to fully decomposed organic matter (Nolan et al.,
59 2011). Therefore, the level of C:N in forest soils is one of the important predictors for estimating
60 soil functions such as C storage capacity of soil and biomass production. When integrated into risk
61 assessment, these functions can serve for modelling scenarios of soil sustainability with climate
62 change issues (Carre et al., 2010). The abovementioned issues highlight that quantifying the

concentration and spatial distribution of SOC, N and specifically their ratio (C:N) are useful for evaluating and improving forest soil management, ecological environment monitoring and climate policy establishment.

The concentrations of SOC, N and consequently C:N level in forest ecosystem are different depending not only on the geographic location but also on soil horizon (Lal, 2008; Brahma et al., 2018) due to different stages of decomposition, erosion and leaching (Gibson et al., 2002; Black et al., 2014). For instance, the decreasing trend of SOC concentration with depth in different forested sites of China and the Czech Republic were reported (Jia et al., 2017; Gholizadeh et al., 2021) and linked to the reduction of below-ground plant biomass with soil depth that as a result has a considerable effect on C:N vertical alteration. Moreover, C:N vertical changes can also happen in response to N leaching (Cools et al., 2014). Lal (2017) stated that there need to be much attention in the storage of SOC in the top 40 cm of soil. Zanella et al. (2017) categorized this depth into organic and mineral horizons. Accordingly, since these soil layers noticeably contribute to concentrations of SOC, N and thus C:N level, particularly in forested areas, the vertical quantification and variability analysis of them should be taken into account.

The number of observations and soil variability to a great extent influence on soil attributes determination at large-scale (e.g., regional and national) studies (Bellamy et al., 2005; Chamberlain et al., 2010). Nevertheless, the cost and efficiency connected to soil sampling, laboratory analysis and assessment of SOC, N and therefore C:N in organic and mineral horizons of forest soils using conventional analytical techniques limit their large-scale prediction due to the requirement of a large number of samples (Vanguelova et al., 2016; Chen et al., 2019), particularly if the aim is measurement of depth (England and Viscarra Rossel, 2018). Quick, cost-effective and reliable approaches for effective determination of SOC, N and C:N in forest soil horizons are thus essential.

Visible–near infrared–shortwave infrared (VNIR–SWIR) spectroscopy is an efficient technique to rapidly and accurately predict soil attributes over large areas at depth, while reducing the cost of soil sampling and analysis (Ben-Dor and Banin, 1995; Gholizadeh et al., 2013). The spectral data acquired from VNIR–SWIR spectroscopy can then sufficiently characterize the spatial (vertical and lateral) as well as the temporal variability of soil (Viscarra Rossel et al., 2017). The technique has been successfully used to measure SOC and N concentrations at different scales (Barthes et al., 2006; Gholizadeh et al., 2011; Knadel et al., 2012; Viscarra Rossel and Hicks, 2015; Gholizadeh et al., 2018). There are also a few local-scale studies studying the use of reflectance spectroscopy for assessing forest SOC in different horizons (Wang et al., 2019; Zhao et al., 2019). Developing large-scale (e.g., national, continental and global) spectral-based model though may be an approach to enhance the success of the technique (Viscarra Rossel et al., 2016). Gholizadeh et al. (2021) used VNIR–SWIR spectroscopy to predict and characterize SOC in different horizons of forest soils over the Czech Republic; however, so far, the capability of VNIR–SWIR spectroscopy for large-scale prediction and spatial analysis of C:N in forest soil layers has not yet fully been exploited.

In this context, the current study attempts to overcome this lacuna and foster VNIR–SWIR reflectance spectroscopy to quantify the levels of SOC, N and C:N in forest soil organic and mineral horizons. In addition, as there is a lack of information on the depth distribution of C:N at the national scale in the Czech Republic, the study aims to evaluate the efficiency of VNIR–SWIR spectroscopy to examine the spatial distribution of C:N in different forest soil layers of the whole country. The premise of this study is that there is a large heterogeneity in the vertical spatial distribution of soil C:N in the forested areas of the country. As VNIR–SWIR spectroscopy can cover large sampling points, both horizontally and vertically, it is also anticipated that the technique

can provide massive SOC, N and thus C:N data, which can facilitate to get a general overview on quality of forest soils in Central Europe.

2. Materials and Methods

2.1. Study sites, soil sampling and laboratory analysis

One-third of the Czech Republic land (78,865 km²) is covered by forests (Czech Statistical Office for Surveying, Mapping and Cadastre, 2019). The sampling sites of the current study were distributed over all the country's forested sites (Figure 1).

Figure 1. Locations of the forests and soil sampling sites in the Czech Republic.

The samples used in this study were originally from the Forest Soil Survey (Fiala et al., 2013). During the years 2000 to 2008, the samples were collected from the forested areas of the whole country. Five-hundred and one (501) samples were separately obtained from three individual organic horizons of litter (L), fragmented (F), humus (H), as well as the mineral horizon of A₁ (with thickness usually from 2 to 10 cm) and the subsurface mineral horizon of A₂ (typically to the depth

of 40 cm) to build a dataset of consisting 2505 samples. The position of each sampling point was recorded using a GeoXM (Trimble Inc., Sunnyvale, CA, USA) receiver with an accuracy of 1 m. Before any analysis, the soil samples were gone through common preparation procedure (air-dried, finely ground, sieved to ≤ 2 mm) and thoroughly mixed (ISO11464:2006). SOC was measured as total oxidized C using wet oxidation (Sparks, 1996) and N was determined by the Kjeldahl digestion distillation method (Bremner and Mulvaney, 1982). The histograms of SOC, N and C:N in different layers can be seen in Figure 2. Since N and C:N in the mineral layers (A_1 and A_2) didn't meet the requirements of a normal distribution, the log transformation of their values was carried out to drastically reduce the skewness and obtain reliable predictions.

SOC N C:N
Figure 2. Histograms of SOC, N and C:N for each soil layer.

2.2. Spectroscopic measurements

Spectral reflectance was recorded across the 350–2500 nm wavelength range using an ASD FieldSpec III Pro FR spectroradiometer (ASD Inc., Denver, Colorado, USA) with a high-intensity contact probe. Figure 3 indicates the scanning setup for the samples' measurement in this study.

Figure 3. Spectra measurement setup.

The spectral resolution of the spectroradiometer was 2 nm for the region of 350–1050 nm and 10 nm for the region of 1050–2500 nm. In addition, the radiometer bandwidth from 350 to 1000 nm was 1.4 nm while it was 2 nm from 1000 to 2500 nm. Based on our laboratory’s protocol, to warm the spectrometer and lamp before any measurement, the instrument turn about 30 minutes earlier on. To avoid beam reflectance from the bottom of the dish (Jensen, 2007), the soil samples were placed in 9 cm diameter petri dishes and formed 2 cm layers of soil. According to Mouazen et al. (2005), a smooth soil surface guarantees maximum light reflection and a high signal to noise ratio (SNR). Thus, to generate a flat surface with the top of the petri dish, the samples were leveled off and spectral scanning was made in three replications (in the center of each sample) in a dark room and the average was saved. The spectroradiometer was optimized using a white Spectralon™ (Lab-sphere, North Sutton, New Hampshire, USA) before the first scan and after every six measurements (Shi et al., 2016).

2.3. Data preprocessing and spectral modelling

All spectra, per soil layer, were preprocessed before modelling. First, we removed the noisy portions between 350 and 400 nm, as well as 2451 and 2500 nm, spectra in the range from 400 to 2450 nm (2051 variables) were kept for further processing. The spectral reflectance (R) was transformed into apparent absorbance spectra through $\log_{10}(1/R)$ and the spectra were then subjected to Savitzky-Golay smoothing (Savitzky and Golay, 1964) with a second-order polynomial fit and 11 smoothing points in order to remove the artificial noise within the working spectral range. The first derivative was afterwards generated and used to remove baseline offset and enhance spectral features (Gholizadeh et al., 2013). Because of high collinearity of the spectra and to reduce dimensionality, 0.1 nm resolution was extracted from whole spectra and we retained only every tenth wavelength from 400 to 2450 nm (Gholizadeh et al., 2021). It is also necessary to

analytically detect reference data outliers in the datasets (Martens and Naes, 1992). The histograms of data, from all soil layers, were therefore plotted and outliers were identified and eliminated, both visually and employing the principle of the Cook distance. This outlier detection technique is applied in regression analysis to detect influential outliers in a set of predictor variables (Cook, 1977; Kim, 2017). The principal component analysis (PCA) was also performed and the scoreplot of the first two principal components (PCs) was used to visualize the structure of the data (Dotto et al., 2018) and to highlight the soil variations in different horizons with different C:N levels.

To build prediction models that given a spectrum will be able to predict either SOC, N and C:N, the data from each soil layer (L, H, F, A₁ and A₂) as well as the dataset obtained from merging all individual layers (profile) were split into training (75%) and testing (25%) datasets using the Kennard Stone (KS) technique (Kennard and Stone, 1969). To develop the regression models and to validate the developed models' generalization capability, the training sets and the testing sets were used, respectively (Kooistra et al., 2003). The spectral modelling was performed using support vector machine regression (SVMR) algorithm with radial basis kernel. The algorithm follows supervised learning based on the statistical learning theory (Vohland et al., 2011). It should be noted that a basic grid search approach was applied to tune SVM's hyperparameters (i.e., cost function and sigma). The implementation of SVM for spectroscopic modelling in detail has previously been described in detail by Viscarra Rossel and Behrens (2010) and Gholizadeh et al. (2016). The spectroscopic models were finally validated using 10-repeated 10-fold cross-validation (Hastie et al., 2009) and the validation statistics provided here are the means of the 10 repeats. The entire spectra preprocessing and modelling procedures were done using the R software (R Development Core Team, Vienna, Austria).

2.4. Prediction performance assessment

The final performance of the models was assessed using standard model evaluation statistics: the coefficient of determination (R^2) values between the observed and predicted values of SOC, N and C:N, Lin's concordance correlation coefficient (LCCC), root mean squared error (RMSE) and bias. R^2 is the proportion of variation in the response that can be described by the regression model and shows the precision of the relationships. RMSE is usually used in VNIR–SWIR spectroscopy to explain the prediction capability of a model and bias is an independent metric that represents the error of means (Bellon Maurel et al., 2010; Gomez et al., 2016), which both indicate the accuracy of the prediction models (Ghebleh et al., 2021). LCCC is a single index that ranges between 0 and 1 (the larger the value, the better the fitting effects) and evaluates the precision and accuracy of the model (Lin, 1989).

$$R^2 = \frac{\sum_{i=1}^n (\text{predicted}_i - \overline{\text{observed}})^2}{\sum_{i=1}^n (\text{observed}_i - \overline{\text{observed}})^2}$$

$$LCCC = \frac{2r\sigma_{\text{observed}}\sigma_{\text{predicted}}}{\sigma_{\text{observed}}^2 + \sigma_{\text{predicted}}^2 + (\overline{\text{predicted}} - \overline{\text{observed}})^2}$$

$$RMSE = \sqrt{\frac{1}{n} \sum_{i=1}^n (\text{observed}_i - \text{predicted}_i)^2}$$

$$\text{bias} = \frac{1}{n} \sum_{i=1}^n (\text{observed}_i - \text{predicted}_i)$$

where predicted_i and observed_i are the predicted and observed values at the time i ; n is the number of total samples; $\overline{\text{predicted}}$ and $\overline{\text{observed}}$ are the mean values of the predicted and observed soil attribute; r is the Pearson correlation coefficient between the predicted and observed soil attribute; $\sigma_{\text{predicted}}$ and σ_{observed} are the standard deviations of predicted and observed soil attribute.

2.5. Spatial distribution of C:N

The spatial distribution of C:N in different forest soil layers acquired through chemical analysis and lab spectroscopy was then mapped as follows:

i) semi-variograms were created to estimate the spatial correlations and the appropriate variogram models.

ii) spatial distribution maps of C:N were obtained using an ordinary kriging method.

Afterwards, the resulting maps were compared to evaluate the capability of prediction models to determine the depth spatial distribution of C:N. The mapping procedure was conducted in the R software (R Development Core Team, Vienna, Austria).

3. Results

3.1. Preliminary findings of soil attributes at different layers

General statistical results of SOC, N and C:N including minimum, maximum, mean, standard deviation (Std.), coefficient of variation (CV) and skewness from all sampling locations for the profile and the individual layers (L, H, F, A₁ and A₂) are shown in Table 1.

Table 1. Statistics description of soil attributes for profile and the individual horizons.

Layer	n	Minimum	Maximum	Mean	Std.	CV	Skewness
SOC (%)							
L	426	36.67	50.91	43.08	2.93	7	0.47
F	431	29.50	48.79	39.46	4.12	10	-0.05
H	446	15.10	42.84	28.80	5.60	19	0.06
A ₁	476	0.82	12.20	5.96	2.29	39	0.02
A ₂	444	0.18	10.70	2.65	1.76	67	1.01
Profile	2223	0.18	50.91	23.29	17.05	73	-0.08
N (%)							
L	426	0.97	2.01	1.44	0.20	14	0.29
F	431	1.03	2.11	1.58	0.20	13	0.27
H	446	0.69	1.86	1.28	0.22	18	0
A ₁	476	0.03	0.55	0.25	0.11	45	0.37

A₂	444	0.01	0.54	0.12	0.08	70	1.28
Profile	2223	0.01	2.11	0.91	0.65	71	-0.15
C:N							
L	426	21.07	47.16	30.54	5.15	17	0.39
F	431	17.69	33.92	25.19	2.90	12	0.08
H	446	14.98	32.10	22.66	3.29	15	0.30
A₁	476	6.16	111.60	26.23	12.20	47	2.84
A₂	444	1.13	112.00	28.83	21.00	73	2.02
Profile	2223	1.13	112.00	26.55	11.77	44	3.43

n: number of samples; Std.: standard deviation; CV: coefficient of variation.

Different layers were remarkably different in case of SOC contents and a large variation in SOC content in soils of the profile as well as the individual layers was seen, which is linked to the different depths from which the soil was sampled. The SOC mean in the L layer was very high (mean = 43.08% $\pm$ 2.93% (Std.)) ranged from 36.67% to 50.91%. These contents decreased to mean = 39.46% $\pm$ 4.12% and 28.80% $\pm$ 5.60% in layers F and H and ranges = 29.50–48.79% and 15.10–42.84%, respectively. In the mineral layers A₁ and A₂, the average SOC contents were noticeably lower (5.96% and 2.65%) than the organic layers (L, F and H) with minimum and maximum of 0.82% and 12.20% for A₁ and 0.18% and 10.70% for A₂. Almost a similar trend can be observed in average of N contents, declining with the depth, except in layer F (mean = 1.58% $\pm$ 0.20%), which showed the highest N contents among all layers (even more than L with mean = 1.44% $\pm$ 0.20%). The N contents were low in mineral layers, particularly in A₂ with minimum and mean values of 0.01% and 0.12%, respectively.

In terms of C:N, the similar pattern of reducing C:N by increasing depth was not followed despite the trend were seen for SOC and N contents. Table 1 indicates that the average C:N in the L layer was higher than all the other individual layers, presenting a high mean value of 30.54 $\pm$ 5.15, while layer H showed the lowest C:N mean of 22.66 $\pm$ 3.29. In addition, there were higher C:N contents in the mineral layers of A₁ and A₂ (mean = 26.23 and 28.83, respectively) compared to the organic layers of F and H. A comparison of the studied layers' CVs highlights that layer A₂ had the highest

variation of C:N ($CV = 73\%$), which shows that its distribution is the most heterogeneous among all soil layers. In contrast, layer F had the $CV = 12\%$, considered as low variability. The data with $CV > 35\%$, $15\% < CV < 35\%$ and $CV < 15\%$ values has been classified as high, moderate and low variability, respectively (Wilding, 1985). The C:N data for all layers positively skewed with the highest skewness of 2.84 in A_1 . The individual organic layer of F had approximated a normal distribution (skewness = 0.08), whereas layers L and H were mildly skewed. The whole profile results highlight that the average C:N of all samples was $26.55\% \pm 11.77\%$, with a distribution that shows rather a moderate variability ($CV = 44\%$) and a highly and positively skewed distribution (skewness = 3.43).

3.2. Soil spectral information at different layers

PCA was applied on the VNIR–SWIR spectra of the forest soil samples from different organic and mineral layers with different concentrations of SOC and N. As shown in Figure 4, some samples were scattered away from the majority of the samples and therefore were considered as outliers. The first two PCs demonstrated about 98.63% variation and the organic layers (L, F and H) spectra were obviously separated from the mineral layers (A_1 and A_2) spectra; however, a spectral similarity was observed between the layers in each group of spectra (Figure 4). This can be related to changes in concentrations and types of the studied soil attributes by depth (Table 1), particularly due to SOC decomposition process (Gholizadeh et al., 2021). The findings of the PCA scoreplot justify the need to investigate the effect of different soil layers on C:N ratio using VNIR–SWIR spectroscopy. It also demonstrates the power of PCA to discriminate VNIR–SWIR reflectance spectra of soils from different depths.

Figure 4. PCA for VNIR–SWIR spectra applied on different soil layers.

Figure 5 presents the average reflectance of soil samples from different layers (L, H, F, A₁ and A₂) and their standard deviation. Average raw spectra shapes and patterns for all soil layers, regardless of the depth, were typical for soil reflectance, with a steady increase through VIS wave range (400–700 nm) and sharp absorption bands, particularly in the SWIR range (1900–2500 nm). The organic layers of L, F and H obviously followed a progression associated with reduction in their organic matter contents; however, the spectra of layer H showed remarkably lower mean reflectance values. Furthermore, the soil samples of the A₁ layer also indicated lower albedo intensity and reflectance across the entire spectra than other the L, F and A₂ layers. Nevertheless, they had wider variance compared to layers L and F. In addition, it can be seen in Figure 5 that the spectra for the forest soil samples collected from the A₂ layer were typical of soils with lower SOC concentration, with common absorptions due to iron oxides, water and clay minerals, mainly in NIR and SWIR regions.

Figure 5. Average reflectance spectra (bold line) and their variance (shaded area) of forest soil samples collected from different layers.

3.3. SOC and N prediction model performance at different soil layers

The results of the SOC and N estimation models at different forest soil layers according to their R^2 , LCCC, RMSE and bias values are presented in Table 2.

Table 2. SOC and N prediction model performance at profile and different soil layers.

Layer	n	R^2	LCCC	RMSE	bias
SOC					
L	426	0.74	0.82	1.75	0.10
F	431	0.77	0.85	1.72	-0.05
H	446	0.63	0.74	3.56	-0.49
A₁	476	0.65	0.74	2.03	0.39
A₂	444	0.71	0.76	1.87	0.38
Profile	2223	0.97	0.99	1.68	0.21
N					
L	426	0.62	0.70	0.09	-0.01
F	431	0.71	0.73	0.07	0

H	446	0.45	0.63	0.18	0.02
A₁	476	0.55	0.66	0.14	0.01
A₂	444	0.60	0.69	0.13	0.01
Profile	2223	0.95	0.98	0.06	0

n: number of samples; R²: coefficient of determination; LCCC: Lin's concordance correlation coefficient; RMSE: root mean square error.

For SOC prediction, the differences between individual layers' models in case of R² were somehow negligible ($0.63 \leq R^2 \leq 0.77$). Layer H showed the lowest R² = 0.63 and also the considerably the highest RMSE and bias values (RMSE = 3.56% and bias = -0.49%) among all individual layers in estimation of SOC, while F performed the best prediction results with the highest R² and lowest RMES and bias (low-biased spatial estimates) values (R² = 0.77, RMSE = 1.72% and bias = -0.05%). In addition, layer F had LCCC = 0.85, which is higher than other layers and shows the best 1:1 fit. The prediction accuracies obtained for assessing N in the individual soil layers in terms of R² were noticeably lower ($0.45 \leq R^2 \leq 0.71$) than those gained for SOC. But similarly, they showed the same trend as SOC, where the F layer (R² = 0.71, LCCC = 0.85, RMSE = 0.07% and bias = 0%), performed better than the other layers and especially than H (R² = 0.45, LCCC = 0.63, RMSE = 0.18% and bias = 0.02%). The most significant finding about N prediction in all examined layers (either individual or merged) was that they were almost unbiased ($-0.01\% \leq \text{bias} \leq 0.02\%$). Furthermore, for both SOC and N, the models developed for the organic horizons of L and F were more accurate than those of the mineral horizons of A₁ and A₂. Table 2 also highlights that for both assessed attributes (SOC and N), the best prediction accuracies with high R² and LCCC and low RMSE values were obtained, when all layers (profile) were used for development of the models (R² = 0.97 and 0.95, LCCC = 0.99 and 0.98, RMSE = 1.68% and 0.06%, respectively). Moreover, the profile models for both attributes showed low error (bias), although the profile model of N was unbiased (bias = 0).

3.4. C:N quantification and spatial distribution at different soil layers

Assessment metrics in Table 3 show the performance of the SVMR models, when predicting C:N in profile and the individual layers. It can be seen that the difference in C:N prediction accuracy between the organic and mineral layers was more considerable than their SOC and N separately. In other words, the assessment metrics obtained for the organic layers in general, and for the L and F data in particular ($R^2 = 0.69$ and 0.71 , LCCC = 0.81 and 0.84 , RMSE = 2.72% and 2.17% and bias = 0.37% and -0.23%), were noticeably better than of the mineral layers of A_1 and A_2 particular ($R^2 = 0.32$ and 0.31 , LCCC = 0.48 and 0.44 , RMSE = 7.66% and 9.52% and bias = 0.85% and 1.26%).

Table 3. C:N prediction model performance at profile and different soil layers.

Layer	n	R^2	LCCC	RMSE	bias
C:N					
L	426	0.69	0.81	2.72	0.37
F	431	0.71	0.84	2.17	-0.23
H	446	0.56	0.66	4.28	0.48
A_1	476	0.32	0.48	7.66	0.85
A_2	444	0.31	0.44	9.52	1.26
Profile	2223	0.63	0.79	7.48	0.46

n: number of samples; R^2 : coefficient of determination; LCCC: Lin's concordance correlation coefficient; RMSE: root mean square error.

According to Table 3, the R^2 and LCCC (0.71 and 0.84) were higher for layer F than other forest soil layers. Likewise, in prediction of samples collected from the layer F, RMSE and bias showed the lowest values (2.17% and -0.23% , respectively) among all examined layers. This was similar to SOC and N prediction results presented in Table 2. The scatterplots of the observed against predicted C:N for each soil layer support these results (Figure 6). Table 3 and Figure 6 also show that merging all individual soil layers, we obtained the soil profile prediction model, which could fairly (Chang et al., 2001) perform ($R^2 = 0.63$, LCCC = 0.79 , RMSE = 7.48% and bias = 0.46%).

The profile model was more accurate than the model for the individual layers of H, A₁ and A₂; nonetheless, less accurate than layers L and F models.

Figure 6. Scatterplots of the observed against predicted C:N at different soil layers.

When the models were validated, the next step was the spatial prediction of C:N in different soil layers using the observed data obtained from wet chemistry and the predicted data derived from VNIR–SWIR spectra. Figure 7 highlights the resulting depth distribution maps, which reflect important information of the soil C:N condition.

It could be observed that the spatial distribution of soil C:N across the forested sites of the country was similar across all soil layers, typically decreasing from west to east. It was also noticed that generally, VNIR–SWIR spectroscopy could fairly predict and spatially analyze C:N in the organic horizons (L, F and H) of the forested sites of the Czech Republic. In these layers, the same tendency

and a coherent spatial distribution were clearly observed between the observed and predicted maps.

Even layer H, with lower accuracy compared to L and F, highlighted the different values of C:N.

Figure 7. Observed and predicted spatial distribution of C:N at different soil layers.

This trend did not work thoroughly well for the mineral layers. For instance, regarding layer A₁, in the easternmost part of the study area, where the lowest values concentrated, a similar spatial trend could be seen between the observed and predicted maps. However, towards the west, C:N pattern and values were quite variable between the maps, without a clearly distinguishable spatial pattern and without depicting the C:N values higher than 50 (Figure 7). The least accurate spatial variability map was for the A₂ layer as the deepest studied layer. It can be seen that similar to layer A₁, high values of C:N (≥ 50) were not detectable in the map created based on the reflectance

spectra and the largest section of the study area was covered with the low and moderate values with no significant spatial variability. It can be mentioned that, in the mineral layers (A_1 and A_2), VNIR–SWIR reflectance spectra were efficient only to map lower values of C:N with highly degraded SOC. In general, findings in Figure 7 highlighted that VNIR–SWIR spectroscopy could detect different classes of C:N in the organic layers, especially L and F, but it failed to differentiate various classes of C:N in forest mineral layers. These results are in agreement with those of presented in Table 3 and Figure 6.

4. Discussion

According to Table 1, the forest soils of the Czech Republic had high SOC concentration in general with the profile mean concentration of 23.29%. In terms of soil layers, the organic layer of L showed the highest SOC (mean = 43.08%), which decreased with increasing soil depth. This trend was proved in previous studies (Ugawa et al., 2012; Gelaw et al., 2013; Zhang et al., 2016, Gholizadeh et al., 2021). This is linked to the litter serving as the main source of forest SOC. The organic composition of litter is gradually decomposed by microorganisms, hence enriching the top soils (Jiang et al., 2017). Moreover, SOC decrease by depth is due to the increasing intensity of microbially-driven decomposition (Kramer et al., 2017). In addition, the most dominant soil class of the Czech Republic and this study's sample set was Cambisols, in which there is little C below the organic horizons. These soils do not show any significant soil development with depth, although they have thick organic rich surface horizons (Deluca and Boisvenue, 2012). In this study, N mirrored the SOC concentrations trend (Table 1) and almost a similar trend was observed between N and forest soil depth, which was also reported in the studies by Kramer et al. (2017) and Sorensen et al. (2020). This can be associated with the mineralization of litter from trees into fine particles

and substances such as ammonium (NH_4^+) that usually sticks to the surface layers (Zhang et al., 2016).

With exception of layer H, all other layers had the values of C:N higher than 24, suggesting that the net nitrification was low (Ollinger et al., 2002; Alberchtova et al., 2008). Nevertheless, high C:N may also be indicative of N immobilization by N restricted soil microbial community (Holub et al., 2005). The organic horizons of L, F and H followed the decreasing trend of C:N, similar to what was observed for SOC and N. Forest floor's (the L layer) C:N ranged from 21.07 to 47.16 (mean = 30.54), which was decreased by depth in the other organic layers (Table 1). This highlights the availability of more amount of fresh organic and less decomposed organic matter in this top layer (Carre et al., 2010). On the other hand, layer H, with the lowest value of C:N among others, represents well-decomposed organic matter and stable humus components. This is attributed to high accumulation of more extensively-decomposed (or microbially-altered) organic compounds in the lower layers after C loss to respiration (Krull and Skjemstad, 2003; Jenkinson and Coleman, 2008). In contrary, it was also clear in Table 1 that in the mineral horizons (A_1 and A_2), the average C:N increased as soil depth increased. This was opposite to the SOC and N trends and the results obtained in a research by Kramer et al. (2017), who found soil C:N values were generally decreasing with increasing of soil depth. Whereas supported by the results yielded in some sites studied by Kramer et al. (2012), Zhang et al., (2016), Wehr et al. (2020) and Radocaj et al. (2012). Working on Hawaiian soils, Kramer et al. (2012), found that organic matter resembling oxidized plant matter accumulated in the deeper soil horizons due to chemical association with short-range order minerals.

The scoreplot of PCA (Figure 4) indicates the data compression and its dimensionality reduction. As expected, there was an obvious separation of data by soil layer (L, F, H, A_1 and A_2). It happened along the continuum of decomposition process and differences in concentrations of soil attributes

(i.e., SOC and N). Based on Figure 4, spectral characteristics were consistent with variation in soil layers. Nevertheless, the obtained outputs were entirely different between organic and mineral layers, which may be linked to the variation in SOC concentrations of the samples that has a direct influence on spectral features and intensity (Ben-Dor and Banin, 1995). Moreover, this can be expected due to significant differences in CV values of SOC, N and C:N between organic and mineral layers (Table 1), which can cause wider scattering of samples in the mineral layers with higher CV values compared to the organic layers.

Figure 5 highlights that the VNIR–SWIR spectra of all soil layers were similar in appearance with respect to pattern, but varied in reflectance values representing variation in N and SOC concentrations and types (Sherman and Waite, 1985; Song et al., 2012). Kuang and Mouazen (2011) and Jia et al. (2014) stated that SOC and N concentrations possess direct spectral response (overtone or combination) in the VNIR–SWIR region. For example, the bands around 1150 nm are attributed to the absorption of amine N–H and the variables near 2450 nm are related to the overtones of C–H bond in methyl (Viscarra Rossel and Behrens, 2010). In the current study, the main factors that dominated the spectral behavior of the soil horizons were probably the wide range of SOC concentrations (0.18%–50.91%) and types (e.g., recognizable fresh remains, humic component). This matches with the conclusions presented by Heil and Schmidhalter (2021). As the layers L and F contain less-decomposed organic matter (e.g., leaves, needles, little branches), their high spectral reflectance is largely caused by remained vegetation photosynthetic pigments (e.g., chlorophyll, carotenoids, xanthophylls) and cell structure (Gitelson et al., 2002). Layer H followed by A₁ indicated the lowest reflectance values compared to the other studied layers (Figure 5), which is probably because of the darker colour of these layers and the well-decomposed structure laying at them (Zanella et al., 2017). However, the high reflectance of the A₂ layer can be associated with

the very low contents of SOC and N in this horizon (Table 1) that was also confirmed by Conforti et al. (2015) and Jiang et al. (2017).

Prediction results in Table 2 indicate the ability of VNIR–SWIR spectra and the models to predict SOC and N on a large scale was satisfactory in forest soils profile (merged individual layers). This may ascribe to strong correlation of SOC and N with soil reflectance as often reported in the literature (Peltre et al., 2011; Dinakaran et al., 2016). The results also show that for both SOC and N, statistics outputs were considerably higher, when all layers (profile) were used for model development. Such results are consistent with those described in studies by Xie et al. (2011) and Jiang et al. (2017), in which the total samples from different depths generally provided better models than the surface and subsurface sets for the measurement of organic matter, SOC and N concentrations, respectively. Kuang and Mouazen (2011) and Jiang et al. (2017) mentioned that the heterogeneity and high CV of the sample sets could increase the accuracy of calibration models. Considering SOC and N prediction in individual horizons (Table 2), it was apparent that the L and F organic layers were predicted more accurately than the mineral layers of A₁ and A₂, which is similar to the results obtained by Jiang et al. (2017). It is probably because the spectral reflectance from the soil, as a predictor, becomes less efficient due to an acute decrease in the SOC and N concentrations (Jia et al., 2017; Gholizadeh et al., 2021). Nevertheless, despite the decline in prediction capability with depth, the VNIR–SWIR spectra could still fairly assess the target attributes in deeper layers of the forested areas across the country. Furthermore, Table 2 indicates that layer F was the best-predicted horizon among the others, which can be associated with the contribution of N and SOC type and their correlation with other soil attributes in this soil layer. This has also been reported in some other studies (Telles et al., 2003; Gholizadeh et al., 2018). In contrast, the H layer had the poorest prediction models of SOC and N that can be linked to its very decomposed materials and high SD values (Table 1) among others (Kuang and Mouazen, 2011; Jia

et al., 2017). In summary, these results revealed successful VNIR–SWIR models for SOC prediction (with high R^2 and LCCC and low error values) and moderate spectral models for N prediction in different forest soil layers, though the N developed models were low- or even zero-biased. These results compared well to previous literature (Ji et al., 2016; Jiang et al., 2017). It should be noted that the soil spectroscopic models established in local-scale studies usually perform better (Stevens et al., 2013; Guerrero et al., 2016), perhaps because the soils sampled at the local-scale have lower variability and more similar spectral features (Gholizadeh et al., 2021).

The findings in Table 3 and Figure 6 explain that C:N prediction using VNIR–SWIR spectra was more accurate in the forest soil profile as well as organic horizons, particularly in the F layer, which is similar to those of obtained for SOC and N individual assessment (Table 2). It is apparent from Table 3 that the developed models for the mineral horizons of A₁ and A₂ were poor and did not have reliable predictive ability. Mutuo et al. (2006) and Heil et al. (2021) were either unable to accurately predict C:N in deeper layers in VNIR–SWIR studies. The very wide range of C:N in A₁ and A₂ layers (6.16–111.60 and 1.13–112.00, respectively) shows the availability of SOC in different decomposition stages, which has different spectral responses and generally causes a drop in model robustness and calibration accuracy due to lack of homogeneity in the population (Brunet et al., 2007; Peltre et al. 2011; Dinakaran et al., 2016). Contrarily, Jiang et al. (2017) showed that the wide concentration range in soil attributes would be advantageous in developing satisfactory VNIR–SWIR spectroscopy models.

The observed and predicted data from wet chemistry and VNIR–SWIR reflectance spectroscopy were used to visualize the depth distribution of C:N in the country's forested sites (Figure 7). The spatial distribution of soil C:N across the forests followed a rather comparable trend across all soil layers reducing from west to east. This might be connected to variation in forest stands (e.g., forest composition, soil classes and altitudes), which affect the SOC, N and C:N dynamics of the forest

soils (Alberchtova et al., 2008; Zhang et al., 2016), particularly in forest floor that is more easily influenced by external factors such as tree species (Ollinger et al., 2002; Zhang et al., 2016). In terms of depth effect, the maps of C:N created by the spectral data in the organic layers (particularly, layer F) generally showed similar spatial distribution patterns with those of obtained from the actual data. The predicted maps in these layers were capable of not only displaying low and medium C:N values, but also detecting high and very high classes of C:N. This can be attributed to rather fair to good prediction accuracy of C:N in organic layers (Table 3 and Figure 6). In the target mineral layers (A₁ and A₂), differences in the range of C:N values occurred and high values were observed, which might be caused by soluble substances (such as NO₃⁻) yielded from litters that are continuously taken into deep soil layers by infiltration (Zhang et al., 2016). However, in spite of the organic layers, VNIR–SWIR spectroscopy was not successful to classify high values of C:N in the mineral layers. This indicates that the effectiveness of C:N spectroscopic models in deeper forest soil horizons was limited. Generally speaking, the current study provided an example on how spectral data can effectively be used for rapid and cost-effective prediction of SOC and N at a national-scale from the upper to lower horizons of heterogeneous forest soils. Furthermore, the obtained outputs highlighted that VNIR–SWIR spectroscopy potential to be a suitable technique for quantifying and mapping forest organic layers (L, F and H) for the purposes of monitoring C:N. Nevertheless, the technique did not support the determination and mapping of C:N in the mineral horizons (A₁ and A₂) of the Czech Republic forested sites.

Detailed C:N information at different depths has the potential to set our understanding of soil functioning (Ben-Dor et al., 2008; Triantafilis et al., 2001). We thus suppose that the study of VNIR–SWIR spectroscopy in C:N assessment at different soil layers will be worthwhile and can further be used to assess soil quality and fertility (Lou et al., 2012; Zhou et al., 2021). It should be stated that there are still some external factors that noticeably affect the value and distribution of

C:N. For instance, tree species and vegetation chemistry (Ollinger et al., 2002) as well as climate regime and topographic factors such as slope and elevation leading to variation in C:N level (Zhang et al., 2016). In addition, Kramer et al. (2012) and Wehr et al. (2020) specified that C:N depth patterns can be driven by soil minerals. Consequently, the capability of VNIR–SWIR spectra for C:N prediction and mapping will be affected, which need to be dealt with in future works.

Conclusions

This study explored the performance of laboratory VNIR–SWIR spectroscopy, SVM machine learning technique and ordinary kriging to quantify SOC and N and consequently to map the spatial distribution of the C:N in different soil layers of the forested sites of the Czech Republic. The conclusions of the study were as follows:

- i) There was a large variation in the studied attributes in forest soil layers depending on the depth from which the samples were taken.
- ii) A similar decreasing trend with depth was seen in average of the SOC and N concentrations, except in the F layer (mean = $1.58\% \pm 0.20\%$), which had the highest N concentration among others. However, for C:N, the similar declining pattern by increasing depth was not observed. Layer L presented the highest C:N (30.54 ± 5.15) though the H layer showed the lowest C:N mean ($22.66\% \pm 3.29\%$).
- iii) Regardless of the depth, the average spectra shapes and patterns for all soil layers were generally similar, although layer H indicated the lowest albedo intensity and reflectance.
- iv) The SOC and N models for soil profile performed satisfactorily ($R^2 = 0.97$ and 0.95 , $LCCC = 0.99$ and 0.98 , $RMSE = 1.68\%$ and 0.06% , bias = 0.21% and 0 , respectively). Moreover, in terms of the prediction accuracy of the individual layers' models, layer F

was the best predicted forest soil layer for both SOC ($R^2 = 0.77$, LCCC = 0.85, RMSE = 1.72% and bias = -0.05%) and N ($R^2 = 0.71$, LCCC = 0.85, RMSE = 0.07% and bias = 0).

v) C:N could be predicted in the soil profile and organic layers, particularly in layer F ($R^2 = 0.71$, LCCC = 0.84, RMSE = 2.17% and bias = -0.23%), but the developed models for the mineral layers poorly performed.

vi) The predicted distribution maps of C:N in different layers of forest soil displayed significant spatial heterogeneity; nevertheless, reflectance spectra could reasonably map the organic layers (especially, L and F).

vii) A noticeable decrease in the map performances of layers A_1 and A_2 was observed, particularly in the areas with high C:N (≥ 50), and it can be stated that VNIR–SWIR spectroscopy failed to show C:N variability in the mineral layers (A_1 and A_2).

viii) Assessment of soil C:N helps to distinguish the SOC decomposition rate in forest soils.

As C:N is very dependent on forest tree species, topographic parameters and soil mineralogy. Further hypothesis testing and evaluating their effects on VNIR–SWIR predictive capability are planned.

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